

Brief Communication Volume <u>2 | Issue 1</u>

# A Concept Note on an in Situ Electrochemical Biosensor and Validation for Metabolites Including Vitamin C and Essential

# Rajib Biswas 1\*, Nirmal Mazumder <sup>2</sup>

<sup>1</sup>Applied Optics and Photonics Laboratory, Department of Physics, Tezpur University, India.

<sup>2</sup>Department of Biophysics, Manipal School of Life Sciences, Manipal Academy of Higher Education (MAHE), Manipal, Karnataka, India-576104.

\*Corresponding Author: Rajib Biswas<sup>1</sup>, Applied Optics and Photonics Laboratory, Department of Physics, Tezpur University, India.

Submitted: 22 Feb 2024 Accepted: 26 Feb 2024 Published: 04 Mar 2024

**Citation:** Rajib Biswas<sup>1</sup>, Nirmal Mazumder<sup>2</sup> (2024). A Concept Note on an in Situ Electrochemical Biosensor and Validation for Metabolites Including Vitamin C and Essential, J of Physics& Chemistry. Reviews & Reports. 2(1). 1-4.

## Abstract

Citrus species possess essential nutritional elements. Being rich in vitamin C, they provide considerable health benefits. Likewise, essential oils of citrus peel vary in quantity as well as quality. This concept-note outlines using a novel electrochemical biosensor for easy and fast detection of citrus metabolites, which will be an immense help in the recognition and segregation of cultivars.

Keywords: Citrus Species, Cultivars, Electrochemical Biosensor.

# Introduction

Citrus species, an important group of fruit crops, are extremely valued for their high vitamin C content and unique, essential oils. Vitamin C content of several Citrus fruits (such as pummelo, Assam lemon, Khasi mandarin, etc.) has been quantified and reported so far in many articles [1-4]. Similarly, the highly acclaimed essential oils of citrus peel also vary in quality and quantity. The commonly used methods for detection of these metabolites are mostly destructive, time- consuming, and laborious. Therefore, the use of sensors for easy and fast detection of citrus metabolites can be of immense help in recognizing and segregating cultivars [5-13]. Therefore, we will use the available collected germplasm to study various aspects like Ascorbic acid content and metabolites, including essential oils. Towards meeting that objective, we propose the following concept note in the form of an electrochemical biosensor which Hyperspectral Imaging further assis

# Design

**Figure 1** shows schematics of the laboratory-based electrochemical biosensing, which will be executed to study the metabolites found in Citrus species. As evident in the scheme, the prototype development is based on two functional units: the first one being the design of the probe cum transducer, which will give the output response while the second one engages the bio functional species, which will be conducive to producing the modulated response. In the execution of the impregnation of bio-functional species, the bio-receptors will be taken in the form of aptamers, enzymes, or proteins, etc., whichever will be better to communicate with the signaling unit. The analyte shown in schematics refers to the processed citrus species. The bio- functional elements will then be characterized through AFM, TEM, FESEM, FTIR etc. to decipher the best binding properties with the target species (analyte).

The other crucial part, i.e., probe cum transducer, will be first simulated for better tunability, and the optimized design will be made ready for impregnation with the tested bio-functional elements.

## **Modulated Response**

Once the transducer is ready as per expediency, the output will be measured through a customized detector, which gives rise to output in the form of current/voltage/power/conductance/resistance. The design of the probe will be first made through COM-SOL, FTTD, MATLAB software. The setup will also provide a wide range of raw data that can be used as per requirement. Accordingly, as per the analyte, the in-situ testing will be executed for metabolites of the citrus species.



Figure 1: Schematic of the Electrochemical Biosensing System.

### Hyperspectral Imaging

**Figure 2:** shows the laboratory-based proposed hyperspectral polarization imaging system, which can aid us in studying the different stages of citrus fruits. The setup is based on an imaging unit and a data processing unit. The polarization measurement will be followed as described in reference 15. The imaging unit consists of a CCD camera and imaging spectrograph. The imaging spectroscope operates in the 400-1000 nm range and will be combined with a CCD camera. This combination will provide a line-scan spectral imaging device. The spectroscope has a spectral resolution of 2.8 nm and a numerical aperture of F/2.4. The CCD camera is monochrome sCMOStype with an 8 mm x 8 mm imaging area, a frame rate of 50 fps, and 16-bit

digital images. The camera setup is mounted to the mounting tower, allowing the alignment of the components in the XZ axis for further adjustment. The lighting assembly consists of halogen lights with a range of ~300-1100 nm and is attached to the mounting tower, which allows height adjustment. The lighting system is connected to the lighting system power supply. The sample is kept on the XY translationstage. The data processing unit consists of a powerful computer with a graphic processing unit. This system will help in fast data acquisition and processing. The advantage of using a custom- made hyperspectral system, other than procuring a hyperspectral camera, is the flexibility of wavelength and setup modification as per the requirement. The setup will also provide a wide range of raw data which can be used as per requirement.

Figure 2: Schematic of Hyperspectral Polarization Imaging System.



**Image Processing** 

Natural pigments in the plants, such as chlorophyll and carotenoids, provide elegant colors to the plants and are responsible for energy production by photosynthesis. Hence, the quantification of pigment levels in the plant can help in grading plant diseases [3-4]. Accordingly, healthy and infected parts from the plant image (including leaves and fruits) could be

segmented using ImageJ software, and different gray level cooccurrence matrix (GLCM) features such as contrast, correlation, energy, entropy etc. are calculated for is segmented region using MATLAB software [6,16-19]. From the GLCM statistics, a significant difference between healthy and infected parts of the plant can be observed, enabling us to decipher the different

#### stages of citrus species.

#### **Concluding Remarks**

In summary, a novel electrochemical biosensor is proposed to estimate vitamin C from different parts of the citrus fruits. It is further asserted that the potential role of the hyperspectral polarizationimaging system will lead to effective assessment of the quality and progression stages of maturation of citrus fruits as well as leaves. It is envisioned that this investigation may help in theidentification of metabolites present and the quality of citrus fruits, which will be a big boon for the agriculture sector.

#### **Funding**

NM thanks the Department of Science and Technology (DST), Government of India, for thefinancial support (project number:

SERB/MTR/2020/000058 and DST/INT/BLG/P-03/2019).

### Acknowledgments

NM thanks Manipal Academy of Higher Education (MAHE), Manipal and Technology Information Forecasting and Assessment Council-Centre of Relevance and Excellence (TIFAC- CORE) in Pharmacogenomics, Manipal School of Life Sciences, MAHE for providing theinfrastructure and facilities.

#### **Conflicts of Interest**

The authors declare no conflict of interest

#### References

- Kakoti R K., Saikia J, Deka S, Gogoi A, Barbora A C (2019) Present Status of Khasi Mandarin in Manipur State of North East India. Int.J.Curr.Microbiol.App.Sci 8: 2157-2165, https://doi.org/10.20546/ijcmas.2019.806.256.
- Bharali R., Bhattacharyya R K, Das P (2017) Bioactive compounds and total antioxidant activity of pummelo (Citrus grandis L.) ecotypes of Assam. Bull. Env. Pharmacol. Life Sci 6: 342-347.
- Feng H, Chen G, Xiong L, Liu Q, Yang W (2017) Accurate digitization of the chlorophyll distribution of individual rice leaves using hyperspectral imaging and an integrated image analysis pipeline. Frontiers in plant science 8: 1238. https:// doi.org/10.3389/fpls.2017.01238.
- Biswas R, Karmakar P K (2020) An Inexpensive and Novel Optical Scheme of Assessing Adulterants in Emulsions, Biointerface Researh in applied Chemistry 10: 6874–6880, https://doi.org/10.33263/BRIAC106.687468805.
- Iwona M., Jamal A, Abderrahim A, Hasnaâ H, Mounsef N, et al. (2021) Assessment of Nutritional, Technological, and Commercial Apricot Quality Criteria of the Moroccan Cultivar "Maoui" Compared to Introduced Spanish Cultivars "Canino" and "Delpatriarca" towards Suitable Valorization. Journal of Food Quality SP - 6679128, https:// doi.org/10.1155/2021/6679128.
- Bhole V, Kumar A (2021) A Transfer Learning-based Approach to Predict the Shelf life of Fruit. Inteligencia Artificial 24: 102–120, https://doi.org/10.4114/intartif.

vol24iss67pp102-120.

- Hamido, Said A. and Morgan, Kelly T. The Effect of Irrigation Rate on the Water Relations of Young Citrus Trees in High-Density Planting. Sustainability 13: 1-18 https://doi.org/ 10.3390/su13041759
- Chen Z, Yu L, Liu W, Zhang J, Wang N (2021) Research progress of fruit color development in apple (Malus domestica Borkh.), Plant Physiology and Biochemistr 162: 267-279, https://doi.org/10.1016/j.plaphy.2021.02.033.
- Mores A, Borrelli GM, Laidò G, Petruzzino G (2021) Molecular Bases of Crop Resistance to Diseases and to Develop Future Breeding Strategies Int. J. Mol. Sci. 22: 5423. https://doi.org/10.3390/ijms22115423
- Yaqoob M, Sharma S, Aggarwal P (2021) Imaging techniques in Agro-industry and their applications, a review. Food Measure 15: 2329–2343 https://doi.org/10.1007/ s11694-021- 00809-w
- Belmonte Sánchez E, Romero González R, Garrido Frenich (2021) A. Applicability of high- resolution NMR in combination with chemometrics for the compositional analysis and quality control of spices and plant-derived condiments, Journal of the Science of Food and Agriculture 101: 3541-3550. https://doi.org/10.1002/jsfa.11051
- Chithra P, Henila M (2021) Apple fruit sorting using novel thresholding and area calculation algorithms. Soft Comput 25: 431–445, https://doi.org/10.1007/s00500-020-05158-2
- García Gómez B E, Salazar J A, Nicolás Almansa, M. Razi, M. Rubio, et al. (2021) M., Ruiz, D., Martínez-Gómez, P. Molecular Bases of Fruit Quality in Prunus Species: An Integrated Genomic, Transcriptomic, and Metabolic Review with a Breeding Perspective. Int. J. Mol. Sci. 22: 333. https://doi.org/10.3390/ijms22010333
- 14. Ancos B D, Rodrigo M J, Sánchez Moreno C Cano, Lorenzo Zacarías (2020) Effect of high- pressure processing applied as pretreatment on carotenoids, flavonoids and vitamin C in juice of the sweet oranges 'Navel' and the red-fleshed 'Cara Cara', Food Research International 132: 109105. https://doi.org/10.1016/j.foodres.2020.109105
- Nirmal Mazumder, Jianjun Qiu, Matthew R Foreman, Carlos Macías Romero, Chih-Wei Hu, et al. (2012) Polarizationresolved second harmonic generation microscopy with a four-channel Stokes- polarimeter. Optics express 20: 14090-9, https://doi.org/10.1364/OE.20.014090
- Sindhoora K.M, Spandana K.U, Ivanov D, Borisova E, U Raghavendra, et al. (2021) Machine-learning-based classification of Stokes- Mueller polarization images for tissue characterization. Journal of Physics: Conference Series 1859 012045 https://doi.org/10.1088/1742-6596/1859/1/012045
- 17. Liu H, runing B, Garnett T, Berger B (2020) Hyperspectral imaging and 3D technologies for plant phenotyping: From satellite to close-range sensing. Computers and Electronics in Agriculture, 175: 105621. https://doi.org/10.1016/j. compag.2020.105621.
- Stien M, Lennart V, Heike S, Kirin D, Katrien, et al.(2021) Proximal Hyperspectral Imaging Detects Diurnal and Drought-Induced Changes in Maize Physiology, Frontiers in Plant Science, 12, 240. https://doi.org/10.3389/

fpls.2021.640914

 Bendel, N., Kicherer, A., Backhaus, A. et al. Evaluating the suitability of hyper- and multispectral imaging to detect foliar symptoms of the grapevine trunk disease Esca in vineyards. Plant Methods 2020 16(142) https://doi. org/10.1186/s13007-020-00685-3

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